

Culture Conditions and ECM Surfaces Utilized for the Investigation of Stem Cell Differentiation

REVIEW / JANUARY 2013

Author: Marguerite Kosovsky, Ph.D.

INTRODUCTION

Stem cells are a natural choice for cellular therapy and regenerative medicine because of their potential for self-renewal and differentiation into a variety of cell lineages. The ability to efficiently drive stem cell differentiation to the lineage of choice will be critical for the success of cellular therapies. To achieve this objective, it will be necessary to investigate the biological properties of diverse stem cell populations, and expand our understanding of the dynamic microenvironment within tissues that governs stem cell behavior.

While great progress has been made in developing methods for expanding undifferentiated stem cells (a promising source for cell-based therapies), there are limited methods currently available for differentiating certain stem cell types in a defined environment. By manipulating the culture conditions in which stem cells differentiate, it has been possible to control differentiation pathways to generate some lineage-specific precursors *in vitro*. This review will focus on the *in vitro* culture systems currently utilized for lineage-specific differentiation of embryonic, induced pluripotent and adult-derived stem cells. Specifically, we will focus on the use of extracellular matrix molecules and a unique peptide-based hydrogel to mimic the stem cell microenvironment and provide conditions that enable a variety of stem cell types to execute specific differentiation pathways.

STEM CELL TYPES

Stem cells are defined by their ability to self-renew (duplication without loss of developmental potential) and to give rise to multiple cell types (differentiation). They can be further defined as pluripotent stem cells and multipotent adult stem cells.

Embryonic stem (ES) cells are derived from the inner cell mass of the embryonic blastocyst. ES cells were first isolated and characterized in mice¹ and subsequently in humans^{2,3,4}. ES cells can differentiate into cells from all three germ layers and give rise to most cell types⁵. The pluripotency of stem cells is controlled by a conserved regulatory network of transcription factors and multiple signaling cascades^{6,7}. These regulatory networks maintain ES cells in a pluripotent and undifferentiated state. Thus, ES cells have great potential for clinical use, but the mechanisms that direct ES cell differentiation *in vitro* and *in vivo* are not well understood. Drawbacks to the clinical use of ES cells are the potential

for immune rejection and ethical controversy surrounding the use of cells extracted from human embryos.

Recent progress with induced pluripotent stem (iPS) cells derived from differentiated cells has potentially expanded the source of therapeutically relevant stem cells. iPS cells are generated by the forced expression of transcription factors such as Oct3/4, Sox2, c-Myc, and KFL-4^{8,9,10,11,12}. iPS cells are thought to be similar to ES cells in many respects, including pluripotency and differentiation potential. Recent work has differentiated both ES and iPS cells under identical methods, on Corning® Matrigel® matrix, laminin, or vitronectin to obtain the same end-product^{13,14}. One notable difference between ES and iPS cells is the cellular epigenetic memory, which may bias the differentiation potential of iPS cells toward lineages of the donor cell^{15,16,17,18}.

CORNING

iPS cells have unique advantages. They are fairly easy to produce, they have been obtained from every human somatic tissue⁵, and they have been made from somatic cells from patients with neurological, metabolic, haematological, cardiovascular and immunodeficiency diseases¹⁷. Thus, iPS cells provide an *in vitro* model suitable for studying specific diseases using patient derived cells and for drug discovery^{19,20,21}. However, there are several barriers to using iPS cells for clinical applications. The first obstacle, which is common to ES cells as well, is the potential for teratoma formation. Even a small number of undifferentiated stem cells can result in germ cell tumors. Thus, a key goal is to differentiate ES or iPS cells to the required cell type as efficiently as possible and minimize or eliminate a residual population of undifferentiated cells¹¹. Secondly, reprogramming of adult somatic cells via viral transfection alters many of the iPS cells to be unstable and thus limits their use. Furthermore, the efficiency of generating iPS cells is low⁷. However, some of these obstacles have been overcome. Recent work has shown that it is feasible to induce iPS cells without viral integration using adenoviruses, miRNAs, or small molecules that demonstrably increase the stability and transduction efficiency of iPS cells^{22,23}.

Adult stem cells are found in adult tissue and organs including the heart, nervous system, gut, and skin. Somatic stem cells are present in most tissues of the adult body, and they play a critical role in response to stress and injury. Adult stem cells are more difficult to study than pluripotent stem cells because they are maintained physiologically in a nearly arrested or non-proliferating state within tissues and organs. Growing them *in vitro* requires them to divide, which is contrary to the quiescent state. Evidence for coexistence of quiescent and active adult stem cells in mammals is found in hair follicle, gut and bone marrow²⁴. A potential disadvantage of adult stem cells is that their ability to differentiate and proliferate decreases with age. They have a more restricted differentiation potential as compared with ES cells²⁵. However, adult stem cells present few ethical concerns and have low immunogenicity²⁶.

The most characterized adult stem cells are blood-derived hematopoietic stem cells (HSC) and mesenchymal stem cells (MSC), which are required to maintain bone, cartilage and other

tissues^{27,28,29}. MSCs are rapidly expanded *in vitro* and sustain differentiation potential to several lineages. The lineage potential and ease of accessibility of MSCs has made them an attractive choice as a cell source for tissue engineering and regenerative medicine³⁰. MSCs have been shown to possess the capability to differentiate *in vitro* into a variety of cell types, including adipocytes, osteoblasts, chondrocytes, myoblasts and neuron-like cells. They have also been shown to differentiate into tissue-specific cells *in vivo* after systematic infusion to treat osteogenesis imperfecta and myocardial infarction^{31,32}. They can be delivered together with various natural and synthetic biomaterials or scaffolds such as tricalcium phosphate and hydroxyapatite ceramics^{33,34}, and are currently utilized in preclinical and clinical studies^{26,35}.

More recently, researchers have reprogrammed adult cells without first passing the cells through a pluripotent state. Fibroblasts have been directly reprogrammed into cardiomyocytes³⁶, blood cell progenitors³⁷ and neurons³⁸. Directed reprogramming may be capable of generating cell-type diversity that is comparable to that derived from pluripotent stem cell intermediates. One unique therapeutic strategy using direct reprogramming is the *in situ* conversion of cell fate. This was demonstrated when pancreatic endocrine cells were converted *in vivo* into insulin producing endocrine cells in the pancreas of a mouse³⁹. A further potential advantage of direct reprogramming is the overall speed and simplicity of the differentiation conditions⁴⁰. Interestingly, reprogrammed cells still need *in vitro* assays that require ECM surfaces to characterize the survival and growth of the newly differentiated cells^{36,37,38,41,42}.

Stem cells have also been isolated from the heterogeneous mixture of cells within tumors and from cancer cell lines. These cancer stem cells (CSC) have the ability to perpetuate through self-renewal and to differentiate to produce diverse mature cell types^{43, 44, 45}. Like stem cells, CSC are controlled by interactions with their microenvironment. Thus, the ECM plays a role in both normal growth and differentiation of stem cells, and in cancer progression^{46,47,48}. For a review on the variety of extracellular matrix substrates utilized in culture systems for growing and studying CSC, see REVIEW, March 2012²⁵¹.

IMPORTANCE OF MICROENVIRONMENT and ECMs for STUDYING STEM CELL DIFFERENTIATION

Classically, the control of stem cell fate has been attributed to genetic and molecular mediators (growth factors, cytokines, and transcription factors). Increasing evidence has revealed that the self-renewal and differentiation of stem cells is controlled by their surrounding environment, which is known as the stem cell niche. In this niche, stem cells communicate with each other via cell junctions, through interactions with the extracellular matrix (ECM), via receptors, and by engaging in dynamic interactions with hormones and soluble factors. The ECM is composed primarily of glycoproteins (laminin, fibronectin, vitronectin), collagens, proteoglycans, and elastin^{49,50}. This material serves to stabilize tissues, promote cell attachment, and modulate cell functionality by specifically interacting with cell surface receptors and activating the associated signaling pathways^{51,52,53}. Integrins are the principal cellular receptors that interact with ECM proteins. ECM proteins

may directly bind specific growth factors and morphogens to integrate signals within the niche microenvironment⁵⁴. The ECM plays structural, biochemical and mechanical roles in normal growth and differentiation of stem cells⁵⁵. The solid state of the ECM, the nanometer scale geometry, matrix elasticity, and the mechanical signals transmitted from the ECM to the cells have been recently shown to direct stem cell differentiation^{30,56,57,220,221,229}.

Recent studies have expanded the range of cell types contributing to the stem cell niche, and include mesenchymal stem cells and macrophage. In mammals, the stem cell niche has been experimentally identified in the bone marrow^{57,58}, nervous system^{60,61}, intestine^{62,63} and skin. While specific components that constitute a particular niche may vary in different tissues, the extrinsic signals that effect self-renewal and differentiation of stem cells appear

to be common, and able to regulate cells at the molecular level through common genetic networks⁶.

Along with the plasticity and multilineage potential of stem cells comes an increased need for regulating their growth and differentiation. However, the mechanisms that enable stem cells to respond with flexibility to their environment have not been well defined^{65,66,67}. Although it is often represented as a static environment, recent studies have demonstrated that the stem cell niche

is spatially and temporally dynamic¹². This allows the stem cells to integrate long term developmental signals with short term cyclical and injury mediated responses. Because of this complexity, the precise and efficient differentiation of stem cells into distinct cell types and tissues is still a major challenge despite rapid advances in stem cell biology. It will be important to better define the interplay of the ECM in the stem cell niche in order to harness the potential of stem cells for treating human disease.

STRATEGIES for STUDING STEM CELL DIFFERENTIATION

Expansion

In order to realize the potential of stem cell therapies, there is a trend to moving toward more defined systems to reduce the risk of contamination from pathogens common to animal sourced materials and to reduce variability and cost^{68,69,70,71,72}. As stem cell research strategies move away from growing stem cells on feeder layers and in non-defined animal-based liquid media, there is also a need to change the culture substrate on which stem cells are grown to make it more chemically defined and less variable⁷³.

The original culture systems designed for embryonic stem cell expansion utilized mouse embryonic feeder (MEF) layers and non-defined media including serum^{1,2,74}. These systems are inefficient for iPSC generation and not applicable to defined, scalable culture for clinical use. Feeder free culture of human embryonic stem cells (hESCs) was first established using Corning® Matrigel® matrix and laminin in conjunction with conditioned media from MEF⁷⁵. Corning Matrigel matrix is a gelatinous, reconstituted basement membrane extract from Engelbreth Holm Swarm mouse sarcoma. More defined protocols with Corning Matrigel matrix were later established to remove conditioned media or serum from the culture systems^{70,76,77}.

Extracellular Matrix proteins (Laminin, collagen IV, vitronectin and fibronectin) were found to support hESC proliferation in the absence of serum⁷⁸ and without Corning Matrigel matrix⁷⁶. Laminin⁷⁹, fibronectin^{80,81}, vitronectin^{82,83}, and laminin/entactin complex⁸⁴ have all been utilized to expand human-derived stem cells. More recently, recombinant Laminin-511 was shown to permit self-renewal of hESCs for at least four months with normal

karyotype and the ability to form cells from all three germ layers⁸⁵. Now, we are realizing that there is a synergistic effect of medium, matrix and exogenous factors on human ESC⁸³.

Li and colleagues were the first to produce a completely synthetic ECM surface⁸⁶. These synthetic polymer matrixes provided xeno-free culture, but only allowed for short term propagation⁸⁷. Using a high throughput screening approach, Brafman et al identified a synthetic polymer that supports the long term self-renewal of pluripotent stem cells⁸⁸. More recently, synthetic peptides have been attached to acrylate surfaces and shown to support proliferation as well as differentiation of hESC into cardiomyocytes⁸⁹. Biomimetic Peptides such as Corning® Purecoat™ ECM Mimetic Cultureware (Fibronectin and Collagen I Mimetic Surfaces) serves as animal-free surfaces for expansion of mesenchymal stem cells⁹⁰ as well as other primary and progenitor cell types. These Biomimetic surfaces contain the active cell binding domains of ECM proteins, synthetically synthesized and covalently bound in a functional orientation.

Finally, synthetic hydrogels supplemented with ECMs or growth factors have been shown to maintain short-term pluripotency and the undifferentiated state of hESC in xeno-free culture systems^{91,92}. However, such hydrogel systems have not been able to maintain the long-term pluripotency of hESCs using xeno-free culture medium. In contrast, Corning Matrigel matrix coated surfaces combined with a chemically defined medium containing xenogenic proteins provides a feeder-free system that is capable of supporting long-term proliferation of hESC⁷⁰.

Differentiation

Traditionally, embryonic stem cells have been induced to differentiate via three fundamental methods, embryoid body (EB) formation, using a feeder layer comprised of stromal cells, or cultured on ECM-based culture substrates^{93,94}. EBs spontaneously differentiate into derivatives of the three embryonic germ layers⁹⁵ upon removal of factors that maintain the undifferentiated state⁹⁶. EBs can be formed using different methods such as dissociated suspension culture, methylcellulose culture, or hanging drop culture^{97,98}. The heterogeneity of EBs and the difficulties associated with producing EBs of uniform size often result in heterogeneous differentiation profiles³.

EBs have been further terminally differentiated using 2D tissue culture plates coated with gelatin followed by supplementation with growth factors and other bioactive factors that induce differentiation. In addition to gelatin, EBs have been differentiated on ECM culture surfaces such as laminin^{15,99}, Corning Matrigel matrix^{100,101,102}, and matrix of human origin from decellularized foreskin fibroblasts¹⁰³. Differentiation of EBs can also occur in 3D hydrogels composed of Collagen or Corning Matrigel matrix¹⁰⁴. Interestingly, the addition of fibronectin to a collagen-based hydrogel was shown to drive the differentiation of the EBs toward endothelial cells, whereas supplementation of collagen with laminin was shown to produce cardiomyocytes¹⁰⁵.

Differentiation

Co-culturing ESC on stromal cells can provide the advantage of growth factors, but these same factors may lead to the differentiation of stem cells to an undesired cell lineage and may also result in contamination from animal-derived components¹⁰⁶. In fact, sialic acid contamination has been found in stem cells grown in the presence of animal products⁶⁸. Recently, researchers have utilized human-derived stromal cells as feeder layers, such as human foreskin fibroblasts, to isolate ESCs under xeno-free culture conditions¹⁰⁷. This strategy does not address other drawbacks when using feeder layers systems for stem cell expansion, including high cost and variability.

Differentiation on monolayers comprised of known ECM-based substrates is one of the simplest protocols. With this method, the type of matrix protein(s) can be well defined. ECM-based surfaces offer considerably more control over the culture microenvironment, and may better mimic the stem cell differentiation niche. In fact, the use of diverse culture surfaces may be an ideal way to promote differentiation into specific lineages. For example, substrates of different shape, nanotopography, or stiffness have been found to direct MSC differentiation into either adipose or osteogenic lineages, even in the absence of biochemical factors^{55,57,108,109,110,111}.

In addition to 2D culture, stem cells can be differentiated in 3D culture environments. Tissue engineering approaches have been used to design synthetic and natural scaffold materials^{112,113}. Considerable research has focused on identifying biomaterials/scaffolds that can provide a 3D culture system that is conducive to stem cell growth and differentiation, and ultimately is capable of mimicking the *in vivo* microenvironment²⁵. A synergistic approach to studying stem cells has brought together cell biologists, biomaterials specialists, and engineers. This interdisciplinary field of stem cell research and bioengineering is focused on creating novel 3D culture systems that promote stem cell differentiation and tissue regeneration. Nanobiomaterials have been developed as substrates for stem cell differentiation^{114,109} and approaches such as electrospinning, micropatterning and microengineered cell adhesive substrates^{114,115} are being utilized to create artificial stem cell niches^{12,61,117}. For these tissue-engineering strategies to be successful, the dynamic relationship between stem cells and the ECM must be understood⁵⁰. This knowledge will enable researchers to more effectively design artificial ECMs that can control stem cell behavior, and further expand our capabilities to produce therapeutically relevant differentiated cell types *in vitro* from adult or pluripotent stem cells³⁰.

DIFFERENTIATION of PLURIPOTENT STEM CELLS

Many laboratories have utilized purified ECM proteins (e.g., laminin, fibronectin, and collagens) or a more physiological material such as Corning® Matrigel® matrix as a surface for pluripotent stem cell differentiation. Other studies have utilized synthetic hydrogels or scaffolds that are functionalized with ECM proteins or peptides. **TABLE 1** provides an overview of culture surfaces utilized for the differentiation of a variety of cell types (and tissues) derived from pluripotent stem cells.

Importance of 3D Differentiation

The optimization of stem cell differentiation protocols have focused primarily on the timing of administration and concentration of growth factors. However, stem cell fate choices are also critically impacted by ECM: stem cell interactions^{30,54,93,97,111}. Although many studies have described ECM preparations, including synthetic surfaces, for the effective attachment and culture of undifferentiated ESCs and iPSCs, the ability of ECM molecules to promote differentiation is less understood. The following two examples highlight the use of Corning Matrigel matrix as a 3D surface for the differentiation of cardiomyocytes, and 3D organogenesis of intestinal crypts.

Recently, Zhang and colleagues have described a novel protocol for the robust differentiation of cardiomyocytes using an overlay system comprised of Corning Matrigel matrix¹⁶⁰. In this protocol, the sequential application of growth factors onto ESC or iPSC cultures in a Corning Matrigel “Matrix Sandwich” generated cardiomyocytes with high purity, and allowed for the epithelial-mesenchymal transition required for the generation of precardiac mesoderm (FIGURE 1A). The mesoderm differentiation of iPSCs grown in the matrix sandwich was determined by examining the expression of Brachyury. Brachyury positive clusters were markedly increased by Corning Matrigel matrix overlays as compared with controls grown without the Corning Matrigel overlay (FIGURE 1B).

Furthermore, the sarcomeric organization of differentiated cardiomyocytes isolated from the matrix sandwich was analyzed by immunolabeling of myofilament proteins alpha-actin and myosin light chain, and confirmed cardiomyocyte differentiation (FIGURE 1C). This novel Matrix Sandwich approach suggests that the addition of ECM in the form of Corning Matrigel matrix provides a favorable microenvironment, complementing the sequential addition of growth factors, to promote a robust differentiation process.

Another example of a study that utilized Corning Matrigel matrix for 3D differentiation of pluripotent stem cells *in vitro* comes from Spense et al¹⁴. These researchers generated a complex 3D organ from ESC (and iPSC) *in vitro* using Corning Matrigel and a temporal series of growth factor manipulations to mimic embryonic intestinal development (FIGURE 2). Figure 2A shows the 3D intestine-like organoids that formed highly convoluted epithelial structures surrounded by mesenchyme after 13 days grown in Corning Matrigel matrix. Figure 2B-E show the expression of intestinal transcription factors after 14 and 28 days of differentiation and reveal that the epithelium matured into columnar epithelium with villus-like involutions that protrude into the lumen of the organoid. This 3D Corning Matrigel culture system has been previously used to differentiate adult stem cells into 3D tissue of the intestine¹⁸⁹, stomach¹⁹⁰ and colon¹⁹¹. In addition, this differentiation system has also been shown to support the differentiation of cancer stem cells¹⁹². Interestingly, recent reports have demonstrated that Corning Matrigel matrix can be used *in vitro* to generate optic cup¹⁶⁹, pituitary¹⁷⁵, and thyroid tissue¹⁷⁴ from embryonic stem cells. Therefore, this differentiation system has broad utility for pluripotent, adult-derived and cancer stem cells.

In addition to Corning Matrigel matrix, individual ECM proteins have been utilized to elucidate the differentiation process with pluripotent stem cells. Battista and colleagues used fibronectin and laminin

interspersed in semi-interpenetrating polymer networks comprised of collagen type I fibers to investigate the influence of the physical and structural properties of the scaffold on mouse ESC differentiation¹⁰⁵. Interestingly, both the composition and strength of the matrix were found to play a role in the formation of EB from ESCs. The presence of fibronectin in the 3D collagen constructs stimulated endothelial cell differentiation and vascularization of the stem cells. On the other hand, the presence of laminin increased the ability of the ESCs to differentiate into beating cardiomyocytes. These findings highlight the importance of scaffold-mediated biological signals in guiding EB differentiation to specific lineages *in vitro*.

A more recent example of ECM proteins used for stem cell differentiation comes from studies generating retinal pigmented epithelium (RPE). Using a novel co-culture system with an

osmolarity gradient, Nistor et al generated 3D retinal progenitor tissue constructs from hESCs¹⁶⁸. Specifically, hESC derived neural retinal progenitors were co-cultured with hESC derived RPE cells on a cell culture insert coated with a 3D mixture of collagen type I and laminin to derive organized retinal tissue. In a separate study, Rowland and colleagues tested various purified mouse and human ECM proteins (laminin-111, laminin-332, fibronectin, collagen, vitronectin, elastin, and Corning® Matrigel® matrix) to generate RPEs from both hESCs and iPSCs¹⁷¹. This work demonstrated that a laminin-111 substrate generated the highest yields of functional RPEs, which was greater than that observed with Corning Matrigel matrix. The identification of a key ECM may help in the future development of a defined, xeno-free, GMP-compliant culture system for the derivation of RPEs that are clinically relevant.

Figure 1. (A) Total cardiomyocytes (CM) present per 35 mm well of the control (without Corning Matrigel overlay) or the matrix sandwich culture for 1-5 days with Activin A (100 ng/mL) added on day 0 followed by bone morphogenetic protein 4 (BMP4) (10 ng/mL) and basic fibroblast growth factor (bFGF) (5 ng/mL) added on days 1 to 5. (B) Fluorescent images of day 2 cells differentiated under control conditions (without Corning Matrigel overlay) or the matrix sandwich culture immunolabeled with Brachyury (Bry) antibody. Scale bars are 200 μ m for the left and middle panels and 50 μ m for the right panel. (C) Epifluorescence images of single CM isolated from the matrix sandwich culture shows sarcomeric organization. Replated CMs from the matrix sandwich culture were colabeled with anti- α -actinin antibody, which marks the Z-lines and anti-myosin light chain (MLC) 2a antibody that shows the A bands in the sarcomere. Scale bars are 20 μ m (Zhang et al 160).

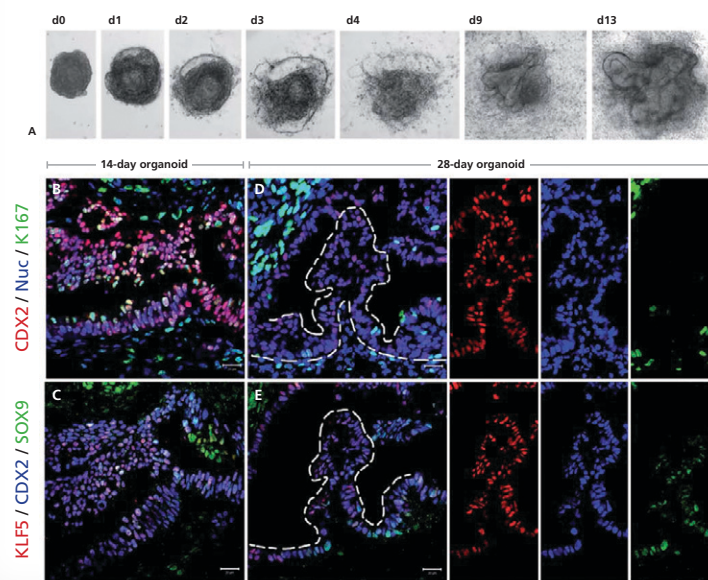
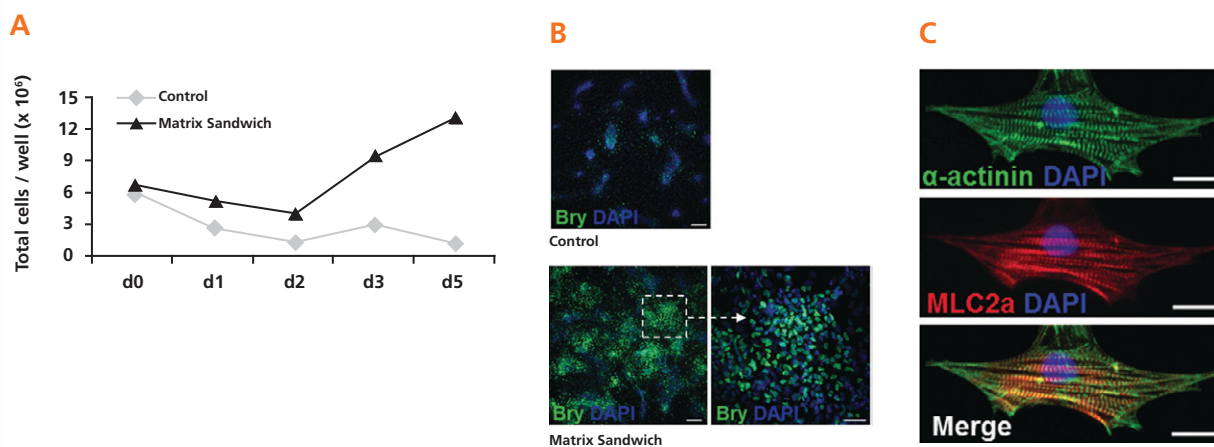


Figure 2. Human ES cells and iPSCs form three-dimensional intestine-like organoids. (A) A time course shows that the intestinal organoids formed highly convoluted epithelial structures surrounded by mesenchyme after 13 days. (B-E) Intestinal transcription factor expression (KLF5, CDX2, SOX9) and cell proliferation on serial sections of organoids after 14 and 28 days (Spence et al 14).

TABLE 1: SURFACES for *in vitro* DIFFERENTIATION of PLURIPOTENT STEM CELLS

Stem Cell Source	Differentiated Cell Type/Tissue	Surface	Reference
mESC,hESC	Germ Cell Layers:ectoderm, mesoderm and endoderm	MEF Feeder Layer	1,2,3,106
hESC	Germ Cell Layers:ectoderm, mesoderm and endoderm	EB-Gelatin	118
mESC	Germ Cell Layers:ectoderm, mesoderm and endoderm	EB-Laminin	99
hiPSC	Germ Cell Layers:ectoderm, mesoderm and endoderm	Corning® Matrigel® Matrix, Vitronectin	119
hpSC	Endoderm	3-D Collagen I Gel	120
hESC	Endoderm	Corning Matrigel Matrix	121
mESC	Mesoderm	Laminin-111	122
mESC,hESC	Mesoderm	Collagen IV	123
hESC	Smooth Muscle	Hydrogels with MEF conditioned media	124
ESC and iPSC	Smooth Muscle	Corning Matrigel Matrix or Feeder Layer	125
hESC	Smooth Muscle	2-D and 3-D Synthetic Fibrous Scaffolds coated with Corning Matrigel Matrix, Laminin, Fibronectin, Vitronectin, or Poly-D-Lysine	126
mESC	Neuronal tissue	Corning PuraMatrix™, Corning Matrigel Matrix, Collagen and Laminin	127
hESC	Neurons	Corning Matrigel Matrix, serum-free	77
hESC and hiPSC	Neuronal tissue	Corning Matrigel Matrix	13
hESC	Neural precursors	Collagen, 2-D and 3-D HA Matrix	128
hESC	Neural precursors	Poly-D-Lysine/Laminin	129, 130
mESC	Neural progenitor cells	Poly-D-Lysine	131
hESC and mESC	cortical neurons, cortical tissue	SFEB Culture	132, 133
mESC	rostral hypothalamic differentiation	SFEB Culture	134
mESC	neurons	3-D Collagen Scaffolds	135, 136
mESC	neurons	PLGA and PLLA Scaffolds coated with Corning Matrigel Matrix or Fibronectin	102
mESC	neurons and astrocytes	EB, 2-D and 3-D Fibrin Scaffold	137
mESC	cortical pyramidal neurons	Gelatin, Poly-D-Lysine	138
rESC	Neural epithelial progenitors	Laminin	139
hESC and hiPSC	Neuronal tissue	EB-Laminin	15
hESC	Neurons	2-D and 3-D Synthetic Fibrous Scaffolds coated with Corning Matrigel Matrix, Laminin, Fibronectin, Vitronectin, or Poly-D-Lysine	126
hESC and iPSC	Osteogenic	Corning Matrigel Matrix	13
hESC	Osteogenic	EB, Poly-D-Lactide scaffold	140
mESC	Osteogenic	Nanofiber Scaffold	141
mESC	Osteogenic	Corning PuraMatrix	142
hESC and hiPSC	Intestinal	Corning Matrigel Matrix	14
mESC	Hematopoietic	Laminin, Corning Matrigel Matrix, Corning Cell-Tak™	143
mESC	Hematopoietic	EB-coculture on OP9 feeder layer	144
mESC	Hematopoietic	3-D tantalum Scaffold	145
hESC	Endothelial	Corning Matrigel Matrix	146, 147
mESC	Endothelial	Collagen Gel with Fibronectin	105
hESC	Endothelial	EB-Corning Matrigel Matrix	101
mESC	Endothelial	Corning Matrigel Matrix	104
hESC and hiPsc	Endothelial	Corning Matrigel Matrix, serum-free	148
mESC	Endothelial	Laminin, Corning Matrigel Matrix, Corning Cell-Tak	143
hESC and hiPSC	Endothelial	Corning Matrigel Matrix, Feeder Layer	125
mESC and miPSC	Endothelial	Corning Matrigel Matrix, Feeder Layer	149
hESC	Cardiomyocytes	Monolayer, Corning Matrigel Matrix	77
hESC	Cardiomyocytes	GF-reduced Corning Matrigel Matrix	150
mESC	Cardiomyocytes	EB-gelatin	151, 152
hESC	Cardiomyocytes	EB-gelatin	151, 153, 154
mESC	Cardiomyocytes	Collagen Gel with Laminin	105
mESC	Cardiomyocytes	EB-Collagen-gel with Corning Matrigel Matrix supplement	155
mESC	Cardiomyocytes	Laminin, Fibronectin, Collagen on microarray	156
Chicken ESC	Cardiomyocytes	Collagen on haluronic acid hydrogels	157
hESC and hiPSC	Cardiomyocytes	Coculture with endodermal cells	158
hESC	Cardiomyocytes	Synthetic peptide-acrylate surface	89
hESC and hiPSC	Cardiomyocytes	Monolayer, Corning Matrigel Matrix	159
hESC and hiPSC	Cardiomyocytes	Corning Matrigel Overlay	160
hESC	Lung Aveolar epithelial	Corning Matrigel Matrix	161
hESC	Lung Aveolar epithelial	EB-gelatin	162

TABLE 1: SURFACES for *in vitro* DIFFERENTIATION of PLURIPOTENT STEM CELLS

Stem Cell Source	Differentiated Cell Type/Tissue	Surface	Reference
mESC	Lung Aveolar epithelial	Decellurized Matrix (mouse lung), Corning® Matrigel®, and Collagen I	249
mESC	Lung Epithelium	EB-gelatin	163
mESC and patient specific iPSC	Respiratory epithelium (Tracheospheres)	ECM Matrix	164
hESC	Retina	Corning Matrigel, Laminin, Feeder layer	165
hESC	Photoreceptors	Corning Matrigel Matrix	166
hESC	Retinal Pigment Epithelium (RPE)	Serum-free Adherent Culture, Corning Matrigel Matrix	167
hESC	3-D Retina	Collagen/Laminin or Corning Matrigel Matrix	168
hESC	3-D Optic Cup	3-D Corning Matrigel Matrix	169
hiPSC	RPE	EB-Gelatin	170
hiPSC	RPE	Corning Matrigel Matrix, Laminin-111	171
hiPSC	Photoreceptors	Corning Matrigel Matrix	166
siPSC	Photoreceptors	EB-Corning Matrigel Matrix, LN, FN	100
hiPSC	RPE	Corning Matrigel Matrix	172
hiPSC	RPE	Serum-free adherent culture, Corning Matrigel Matrix	167
hESC	Thyroid	3-D Corning Matrigel Matrix	173
mESC	Thyroid epithelium	EB-gelatin	163
mESC	Thyroid Follicular Cells	3-D Corning Matrigel Matrix	174
mESC	Adenohypophysis tissue (Pituitary)	Aggregate culture	175
hESC	Hepatic progenitor cells, Cholangiocyte like cells	Collagen I, Corning Matrigel Matrix	177
hESC	Biliary lineage	Collagen I	177
hESC	Hepatic endoderm	Laminin, Collagen I, Corning Matrigel Matrix	178
rESC	Hepatocyte	Corning Matrigel Matrix	179
hiPSC	Hepatic endoderm	Corning Matrigel Matrix	180
hiPSC	Hepatocyte	Corning Matrigel Matrix	181
hiPSC	Hepatocyte	GF-reduced Corning Matrigel Matrix	18
mESC	Hepatocyte-like cells	EB-Gelatin	182
hESC	Hepatocyte-like cells	Corning Matrigel, conditioned media from HepG2-CM	183
hESC	Hepatocyte-like cells	2-D and 3-D Synthetic Fibrous Scaffolds coated with Corning Matrigel Matrix, Laminin, Fibronectin, Vitronectin, or Poly-D-Lysine	126
hESC	Pancreatic exocrine cells	EB-gelatin	184
hESC	Pancreatic endoderm	Corning Matrigel Matrix	77
hESC	Pancreatic progenitors	Feeder Layer	185
hESC, hiPSC	Insulin producing pancreatic cells	Corning Matrigel Matrix	186
hESC	Adipose	Human Foreskin Fibroblast ECM Extract	103
mESC	Chondrocytic	Alginate encapsulated EB	187
mESC	Chondrocytic	2-D and 3-D Scaffolds of Polyethylene glycol	188
mESC	Chondrocytic	Cartilage extract	104

DIFFERENTIATION OF Adult Stem Cells

In contrast to ESC and iPSCs, there have been no reports to date of adult stem cells differentiating into tumorigenic cells. Consequently, adult-derived stem cells are considered to be a very promising source of cells for tissue engineering and cell-based therapies⁷. Similar to the work described above for pluripotent stem cells, many different ECM surfaces have been utilized to investigate the differentiation of adult stem cells. Both naturally occurring and synthetic molecules have been used. **TABLE 2** provides an overview of the culture surfaces utilized for the differentiation of a variety of cell types (and tissues) derived from adult stem cells.

Physical and Mechanical Cues for Differentiation

Stem cell differentiation has become increasingly linked to mechanobiological concepts such as cell generated physical force and nanotopological structure. By examining the differentiation behavior of MSCs cultured *in vitro* on different substrates, scientists are beginning to understand the dynamic physical and mechanical properties that control the stem cell niche.

Kilian et al have elegantly demonstrated the effect of cell shape on MSC differentiation. This study used fibronectin as a coating on synthetic surfaces of different geometries constructed using a microcontact printing strategy⁵⁷. The shape of the individual synthetic substrate, either star shaped or flower shaped, strongly influenced the cytoskeletal organization within these adherent cells (FIGURE 3A-E). Furthermore, the star shape directed

differentiation into osteoblasts while the flower shape directed differentiation into adipocytes (FIGURE 3F). This work established that even subtle shape changes play a significant role in promoting stem cell differentiation. Furthermore, it highlights the critical need for functionalization with ECM or peptides for optimal behavior of the synthetic substrates. Similarly, polyacrylamide gels of varying stiffness were functionalized with collagen and found to drive the differentiation of MSCs to muscle, bone or neuronal lineage depending on the stiffness of the substrate^{108,239}, and PEG hydrogels of varying stiffness functionalized with laminin affected adult mouse stem cell differentiation⁵⁶. These findings are further supported by recent differentiation studies of MSCs on nanotubes of differing diameters¹⁰⁹. Oh and colleagues found that as the nanotube diameter increased, there was an increase in MSC differentiation into osteoblastic lineages.

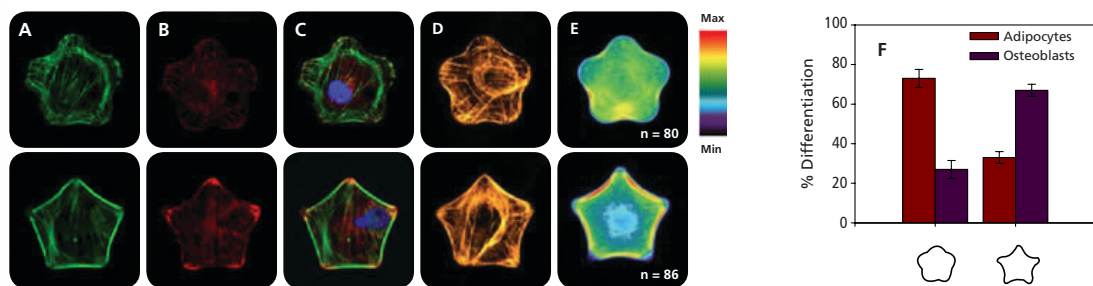


Figure 3 A-E. (A-C) Immunofluorescent images of cells in flower and star shapes stained for F-actin (green), vinculin (red) and nuclei (blue). (D) Immunofluorescent images of cells in flower and star shapes stained for myosin IIa. (E) Fluorescent heatmaps of >80 cells stained for myosin IIa as a quantitative measure of contractility (Scale bar, 20 microns). (F) Percentage of cells differentiating to adipocytes or osteoblasts when cultured on patterns of the same shape. p- value < 0.01 (Kilian et al 57).

TABLE 2: SURFACES for *in vitro* DIFFERENTIATION OF ADULT STEM CELLS

Stem/Progenitor Cell Type	Differentiated State	Extracellular Matrix or other Surface for Differentiation	Reference
INTESTINAL	Intestinal organoids	Corning® Matrigel® Matrix	193
	Crypt villus structure	Corning Matrigel Matrix	189, 194
	Crypt villus structure	Collagen	195
COLON	Epithelial organoids	Corning Matrigel Matrix	191, 196
	Epithelial organoids	Collagen I	197
STOMACH	Gastric Units	Corning Matrigel Matrix	190
MAMMARY	Functional mammary glands	Corning Matrigel Matrix	198, 199, 200
EPIDERMAL	Keratinocytes	Fibroblast feeder layer	201
	Keratinocytes	Fibronectin, Collagen I, Laminin on micropatterned surfaces	202
HAIR FOLLICLE	Neurons, smooth muscle	Corning Matrigel Matrix	203
	Melanocytes	Fibronectin	203
PANCREATIC	Endocrine cells	Corning Matrigel Matrix	204
	Endothelial	Laminin, Collagen IV, Netrin 1, Netrin 4	205
	Pancreatic endocrine and exocrine cells, neurons and glial cells, stellate cells	Corning Matrigel Matrix	206
MUSCLE	Skeletal muscle	Laminin covalently attached to PEG Hydrogel with different stiffness	56
CARDIAC PROGENITOR	Cardiomyocyte	Corning Matrigel Matrix, Collagen I, and Poly-D-Lysine	207
	Cardiomyocyte	Collagen coated HA Hydrogels of varying stiffness	157
ADIPOSE	Endothelial	Corning Matrigel Matrix	209, 210
HEPATIC OVAL	Pancreatic	Serum-free Suspension	211
HEPATIC PROGENITOR	Hepatic or biliary cells	Laminin, Synthetic polyurathane	213
	Endothelial cells and osteocytes	Collagen	212
LUNG	Bronchioalveolar epithelium	Corning Matrigel Matrix	214
SALIVARY GLAND	Salivary epithelium	Corning Matrigel Matrix, Corning PuraMatrix™	215
RETINAL	Retinal neuron	Corning PuraMatrix	216
	Rod-like cells	Retinal Neurospheres-ECM	217
NEURONAL	Neuron, neuroblasts	Neurospheres, Poly-L-Ornathine, Laminin	218
NEURAL PROGENITOR	Neuron, astrocyte	Neurospheres, Poly-D-Lysine/Laminin	220
	Neuron	IKVAV peptide linked to nanofibrils	223
	Neuron	Corning PuraMatrix + Laminin	226
	Neuron	Corning PuraMatrix	228
NEURAL STEM CELLS	Neuron	3-D culture on Alignate hydrogels	220
	Neuron	Corning PuraMatrix	224
	Neuron	Corning PuraMatrix, Corning Matrigel Matrix	225
	Neuron	Diff in 2-D and 3-D self assembling peptide hydrogels	227
	Neuron, glial and astrocyte	Hydrogel with RGD peptides, Variable moduli of the interpenetrating polymer networks (vmIPNs)	229
PRENATAL RAT CELLS	Neuron and glial cells	Laminin attached to acrylamide gels of differing stiffness	221
MSC	Cardiomyocyte	Collagen IV	231
	Cardiomyocyte	Collagen I linked to polyacrylamide gels of varying stiffness	108
	Cardiomyocyte	Corning Matrigel Matrix, collagen, laminin, fibronectin	250
	Cardiomyocyte	Corning Matrigel Matrix	232
	Cardiomyocyte	Fibronectin on PLGA Thin Film	233
	Cardiomyocyte	Hydrogel + Laminin	234
	Chondrocyte	Fibronectin	235
	Chondrocyte	Corning PuraMatrix	236
	Osteocyte	Corning PuraMatrix	237
	Osteocyte	TiO2 nanotubes	108
	Osteocyte	Corning PuraMatrix	238
	Osteocyte	Collagen I linked to polyacrylamide gels of varying stiffness	108
	Adipocyte	Collagen modified PDMS and polyacrylamide gels	239
	Adipocyte	Corning Matrigel Matrix	209
	Osteocyte, adipocyte	ECM peptides conjugated to synthetic polymers	240
	Osteocyte, adipocyte	FN printed onto PDMS substrate	57
	Hematopoetic	Co-culture with HSCs	241
	Hematopoetic	Corning PuraMatrix	242
	Neuron	Collagen , LM, FN	230
	Neuron	Corning Matrigel Matrix, Fibronectin, Laminin, Collagen	222
	Neuron	Collagen I linked to polyacrylamide gels of varying stiffness	108, 110
	Hepatocyte	FN, Corning Matrigel Matrix	243
	MSC + ENDOTHELIAL PROGENITORS	Microvascular endothelial network	

Nanobiomaterials

Native ECM is constructed through self-assembly of many nanofibrillar proteins secreted by cells. Thus, the normal cell environment is comprised of a network of ECM molecules with nano- and micro-scale architecture. Although many synthetic hydrogels are successful at providing 3D support for stem cells, most fail to mimic the native dimensionality and functional diversity of the ECM environment.

A peptide-based hydrogel has been developed to study stem cells in a defined and 3D environment, which has been shown to support the growth and differentiation of a variety of cell types when supplemented with known bioactive molecules (e.g., growth factors, ECMs)^{224,245,246,247}. This material, Corning® PuraMatrix™ Peptide Hydrogel, contains a synthetic, self-assembling peptide that exhibits a nanometer scale fibrous structure under physiological conditions (average pore size of 50-200 nm). The peptide component is comprised of 16 amino acids with a repeating sequence of Arg, Ala, Asp and Ala [(RADA)₄ or RAD16] and the complete material is composed of 1% peptide and 99% water²⁴⁵. Corning PuraMatrix gives rise to the spontaneous assembly of a water-soluble beta sheet bilayer structure, providing cells with an attachment substrate that exhibits a structure that is analogous to that exhibited by proteins.

While Corning PuraMatrix has been shown to support cell proliferation, attachment, and neurite outgrowth using established cell lines or primary cell types²⁴⁸, more recently it has also been used as a substrate for adult stem cell differentiation of neuronal precursors and MSCs. Thornhoff et al, 2008 found that Corning PuraMatrix was the most optimal hydrogel for human fetal neural stem cells, supporting both cell migration and neuronal differentiation. These cells were isolated and cultured as neurospheres, primed with poly-D-lysine and then mixed with different hydrogels including Corning PuraMatrix and Corning Matrigel® matrix²²⁵. A neural stem cell-like cell type, Muller Glia of the retina, has been shown to differentiate on Corning PuraMatrix²¹⁶. Muller cells exhibited differential expression of neuronal genes when cultured in a 3D system comprised of Corning PuraMatrix, in contrast to cells cultured in 2D on uncoated plates under identical media and growth factor supplementation conditions.

More recently, Ortinau and colleagues have demonstrated the direct influence of a 3D Corning PuraMatrix-based scaffold on the neuronal differentiation of human neural progenitor cells²²⁶. Cells were grown in Corning PuraMatrix supplemented with laminin (PML 0.25 %). Using transmission light microscopy (FIGURE 4A + B) and scanning electron microscopy (FIGURE 4C + D) to follow the induction of differentiation, these researchers could directly show that neural progenitor cells started to develop a dense network of neuronal processes (FIGURE 4B + D). Immunocytochemistry revealed that after 7 days of differentiation in the Corning PuraMatrix-laminin scaffold, the neuronal precursor cells began to express β III-tubulin and tyrosine hydroxylase (TH) (FIGURE 4E + F). They concluded that functionalized Corning PuraMatrix peptide hydrogel may be a valuable tool for the generation of defined cell types, and could be utilized for *in vitro* assays of stem and progenitor cell differentiation.

Moreover, Hamada et al have evaluated the osteogenic differentiation of MSCs, and analyzed the spatial distribution of mineralized bone in 3D using Corning PuraMatrix²³⁷. Their results were confirmed by biochemical, gene expression and microscopic analyses. In addition to *in vitro* studies, recent work has characterized *in vivo* MSC differentiation into osteoblasts using Corning PuraMatrix. Furthermore, the Corning PuraMatrix complexes were implanted subcutaneously into rats and demonstrated feasibility for osteogenesis in an animal model system²³⁸.

Corning PuraMatrix has also been used to mimic the functional hematopoietic stem cell niche *in vitro* by co-culturing human mesenchymal stromal cells (MSCs) with human bone marrow derived CD34+ cells²⁴². Instead of artificially tailoring the composition of the matrix with the addition of purified ECM molecules, the researchers formulated the matrix naturally by culturing MSCs in Corning PuraMatrix. When grown with the Corning PuraMatrix hydrogel, the MSCs were found to secrete high levels of ECM proteins, which interacted with matrix fibers and formed a meshwork-like structure similar to the *in vivo* marrow microenvironment.

Moreover, recent studies have evaluated whether human MSCs and human endothelial precursors can form microvascular networks on scaffolds composed of collagen type I, fibronectin, or Corning PuraMatrix *in vivo*. Corning Matrigel matrix was used as a control scaffold based on previous *in vitro* and *in vivo* studies. The results demonstrated that vascular density was greatest in more compliant ECM scaffolds comprised of Corning PuraMatrix (5 Pa) and Corning Matrigel (80 Pa), as compared to stiffer substrates composed of collagen and fibronectin (385-510 Pa)²⁴⁴. These findings are consistent with related studies, which show that substrate stiffness directly influences stem cell differentiation *in vitro*^{57,108}.

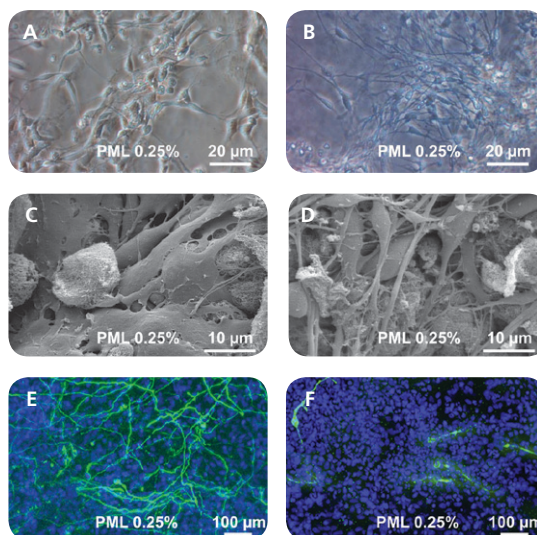


Figure 4. Proliferation and differentiation in 3D scaffold. (A + B) Transmission light picture of proliferating cells in PuraMatrix with 0.25 % laminin (PML) and differentiating cells in PML 0.25% (C + D). Scanning electron microscope picture of proliferating cells and differentiated cells in PML 0.25%. (B + D) Upon induction of differentiation one observes the development of a dense 3 dimensional network of processes. (E) Immunocytochemistry for β III-tubulin and TH of uninduced cells in PML 0.25% and (F) Cells after 7 days of differentiation revealed a dense network of β III-tubulin positive cells. TH+ cells were found to possess processes, but without building up a dense network (Ortinau et al 226).

FUTURE DIRECTIONS

The multilineage potential of pluripotent and adult-derived stem cells presents a challenge as well as an opportunity. Differentiation at the wrong time or in the wrong place or to an undesirable cell type may lead to pathophysiological conditions. Ultimately, scalable animal component-free cell culture surfaces will be essential for the translation of basic research into the clinic. However, these defined surfaces must allow not only for expansion, but also directed differentiation. The convergence of two important disciplines, biomaterials engineering and stem cell research, will allow us to better mimic the physiological complexity of the stem cell niche and ultimately provide the multitude of required cell types for clinical therapies in humans.

REFERENCES

Author information: Marguerite Kosovsky, PhD, is a freelance writer based in Northborough, MA

For further information, please contact Corning Technical Support at 800.492.1110 or email CLSTechServ@corning.com

1. Evans, M.J. and Kaufman, M.H. (1981) Establishment in culture of pluripotent cells from mouse embryos, *Nature* 292:154-156.
2. Thomson, J. A. et al (1989) Embryonic stem cell lines derived from human blastocysts, *Science* 282:1145-1147.
3. Rubinoff, B.E. et al (2000) Embryonic stem cells from human blastocysts: Somatic differentiation in vitro, *Nat. Biotechnol.* 18:399-404.
4. Cowan, C.A. et al (2004) Derivation of embryonic stem cell lines from human blastocysts, *New Eng. J. Med* 350:1353-1356.
5. Alvarez, C.V. (2012) Defining Stem Cell Types: understanding the therapeutic potential of ESC, ASC, and IPS, *J. Mol. Endocrinol* 49:R89-R111.
6. Pera and Tam (2010) Extrinsic regulation of pluripotent stem cells, *Nature* 465:713-720.
7. Higuchi, A. et al (2012) Biomimetic cell culture proteins as extracellular matrices for stem cell differentiation, *Chemical Reviews* 112:4507-4540.
8. Takahashi, K. and Yamanaka, S. (2006) Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors, *Cell* 126:663-676.
9. Takahashi, K. et al (2007) Induced pluripotent stem cell lines derived from human somatic cells, *Science* 318:1917-1920.
10. Jaenisch, R. and Young, R. (2008) Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming, *Cell* 132:567-582.
11. Yamanaka (2009) A fresh look at iPS cells, *Cell* 137:13-15.
12. Wilmat, I. et al (2011) The evolving biology of cell reprogramming, *Phil. Trans. Soc. B* 366:2183-2197.
13. Chambers, S.M. et al (2009) Highly efficient neural conversion of human ES and iPS cells by dual inhibition of SMAD signaling, *Nat. Biotechnol.* 27:275-280.
14. Spence, J.R. et al (2011) Directed differentiation of human pluripotent stem cells into intestinal tissue in vitro, *Nature* 470:105-109.
15. Hu, B.-Y. et al (2010) Neural differentiation of human induced pluripotent stem cells follows developmental principles but with variable potency, *PNAS* 107:4335-4340.
16. Kim, K. et al (2012) Donor cell type can influence the epigenome and differentiation potential of human induced pluripotent stem cells, *Nat. Biotechnol.* 29:1117-1119.
17. Robinton, D.A. and Daley, G.Q. (2012) The promise of induced pluripotent stem cells in research and therapy, *Nature* 481:295-305.
18. Kajiwara, M. et al (2012) Donor-dependent variations in hepatic differentiation from human-induced pluripotent stem cells, *PNAS* 109:12538-12543.
19. Yamanaka, S (2007) Strategies and new developments in the generation of patient-specific pluripotent stem cells, *Cell Stem Cell* 1:39-49.
20. Park, I.H. et al (2008) Disease specific induced pluripotent stem cells, *Cell* 134:877-886.
21. Itzhaki, I. et al (2011) Modeling the long QT syndrome with induced pluripotent stem cells, *Nature* 471:225-229.
22. Anokye-Danso, F. et al (2011) Highly efficient miRNA-mediated reprogramming of mouse and human somatic cells to pluripotency, *Stem Cell Stem* 8:376-388.
23. Valamehr, B. et al (2012) A novel platform to enable the high-throughput derivation and characterization of feeder-free human iPSCs, *Scientific Reports* 2,213; DOI:10.1038/srep00213.
24. Li, L. and Clevers, H. (2010) Coexistence of quiescent and active adult stem cells in mammals, *Science* 327:542-545.
25. Fu, R.-H. et al (2011) Differentiation of stem cells: Strategies for modifying surface biomaterials, *Cell Transplantation* 20:37-47.
26. Si, Y.-L. et al (2011) MSCs: Biological characteristics, clinical applications and their outstanding concerns, *Aging Research Reviews* 10:93-103.
27. Pittenger, M.F. et al (1999) Multilineage potential of adult human mesenchymal stem cells, *Science* 284:143-147.
28. Jiang, Y. et al (2002) Pluripotency of mesenchymal stem cells derived from adult marrow, *Nature* 418:41-49.
29. Lee, (2004) Isolation of multipotent mesenchymal stem cells from umbilical cord blood, *Blood* 103:1669-1675.
30. Guilak, F. et al (2009) Control of stem cell fate by physical interactions with the extracellular matrix, *Cell Stem Cell* 5(1):17-26.
31. Hao, L. (2012) Mesenchymal stromal cells for cell therapy: besides supporting hematopoiesis, *Int. J. Hematol.* 95:34-46.
32. Jackson, W.M. et al (2012) Concise review: clinical translation of wound healing therapies based on mesenchymal stem cells, *Stem Cells Translational Medicine* 1:44-50.
33. Dennis, J.E. (1998) In vivo osteogenesis assay: a rapid method for quantitative analysis, *Biomaterials* 19:1323-1328.
34. Ohgushi, H. (2005) Tissue engineered ceramic artificial joint-ex vivo osteogenic differentiation of patient mesenchymal cells on total ankle joints for treatment of osteoarthritis, *Biomaterials* 26:4654-4661.
35. Ren, G. et al (2012) Concise review: Mesenchymal stem cells and translational medicine: emerging issues, *Stem Cells Translational Medicine* 1:51-58.
36. Leda, M. et al (2010) Direct reprogramming of fibroblasts into functional cardiomyocytes by defined factors, *Cell* 142:375-386.
37. Szabo, E. et al (2010) Direct conversion of human fibroblasts to multilineage blood progenitors, *Nature* 468:521-526.
38. Vierbuchen, T. et al (2010) Direct conversion of fibroblasts to functional neurons by defined factors, *Nature* 463:1035-1041.
39. Zhou, Q. et al (2008) In vivo reprogramming of adult pancreatic exocrine cells to beta-cells, *Nature* 455:627-632.
40. Chambers, S.M. and L. Studer (2011) Cell fate plug and play: direct reprogramming and induced pluripotency, *Cell* 145:827-830.
41. Kim, J. et al (2011) Direct reprogramming of mouse fibroblasts to neural progenitors, *PNAS* 108:7838-7843.
42. Ahmed, R.P.H. et al (2011) reprogramming of skeletal myoblasts for induction of pluripotency for tumor-free cardiomyogenesis in the infarcted heart, *Circulation Res.* 109:60-70.
43. Reya, T et al (2001) Stem cells, cancer and cancer stem cells, *Nature* 414:105-111.
44. Alison, M.R. et al (2010) Stem cells in cancer: instigators and propagators? *J. Cell Science* 123:2357-2368.
45. Visvader, J.E. (2011) Cells of origin in cancer, *Nature* 469:314-322.
46. Borovski, T. et al (2011) Cancer stem cell niche: The place to be, *Cancer Res.* 71:634-639.
47. Leventhal, K.R. et al (2009) Matrix crosslinking forces tumor progression by enhancing integrin signaling, *Cell* 139:891-906.
48. Bissell, M.J. and Hines, W.C. (2011) Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression, *Nature Medicine* 17:320-329.
49. Hynes, O (2009) The extracellular matrix: not just pretty fibrils, *Science* 326:1216-1219.
50. Daley, W.P. et al (2007) Extracellular Matrix dynamics in development and regenerative medicine, *J. Cell Science* 121:255-264.
51. Discher, D.E. et al (2006) Tissue cells feel and respond to the stiffness of their substrate, *Science* 310:1139-1143.
52. Discher, D.E. et al (2009) Growth factors, matrices and forces combine and control stem cells, *Science* 324:1673-1677.
53. Inglesias-Bartolome, R. and J.S. Gutkind (2011) Signaling circuitries controlling stem cell fate: to be or not to be, *J. Cell Science* 23:716-723.
54. Brizzi, M. F. (2010) Extracellular matrix, integrins, and growth factors as tailors of the stem cell niche, *Curr. Opinion in Cell Biology* 24:1-7.
55. Reilly, G.C. and Engler, A.J. (2010) Intrinsic extracellular matrix properties regulate stem cell differentiation, *J. Biomechanics* 43:55-62.
56. Gilbert, P.A. et al (2010) Substrate elasticity regulates skeletal muscle stem cell self-renewal in culture, *Science* 329:1078-1081.
57. Kilian, K.A. et al (2010) Geometric cues for directing the differentiation of mesenchymal stem cells, *PNAS* 107:4872-4877.
58. Di Maggio, N. et al (2011) Toward modeling the bone marrow niche using scaffold based 3D culture systems, *Biomaterials* 32:321-329.

REFERENCES

59. Wagners, A.J. (2012) The stem cell niche in regenerative medicine, *Cell Stem Cell* 10:362-369.
60. Solozobova, V. et al (2012) Lessons from the embryonic neural stem cell niche for neural lineage differentiation of pluripotent stem cells, *Stem Cell Rev and Rep* 8:813-829.
61. Lampe, K.J. and Hen, S.C. (2012) Building stem cell niches from the molecule up through engineered peptide materials, *Neuroscience Letters* 519:138-146.
62. Barker et al (2007) Identification of stem cells in small intestine and colon by marker gene *Lgr5*, *Nature* 449:1003-1008.
63. Medema, J.P. and Vermeulen, L. (2011) Microenvironmental regulation of stem cells in intestinal homeostasis and cancer, *Nature* 474:318-326.
64. Wong, V.W. et al (2012) Stem cell niches for skin regeneration, *Int. J. Biomaterials* ID#926059.
65. Lander, A.D. (2009) The 'stem cell' concept: is it holding us back?, *J. Biology* 8:70.
66. Lander, A.D. et al (2012) What does the concept of the stem cell niches really mean today?, *BMC Biology* 10:19.
67. Ehninger, A. and Trumpp, A. (2011) The bone marrow stem cell niches grows up: mesenchymal stem cells and macrophages move in, *J. Exp. Med.* 208:421-428.
68. Martin, M. J. (2005) Human embryonic stem cells express an immunogenic nonhuman sialic acid, *Nature Med.* 11:228-232.
69. Fink, D.W. (2009) FDA regulation of stem cell-based products, *Science* 324:1662-1663.
70. Higuchi, A. et al (2011) Biomaterials for the feeder-free culture of human embryonic cells and induced pluripotent stem cells, *Chemical Rev.* 111:3021-3035.
71. Weber, D.J. (2006) Manufacturing considerations for clinical uses of therapies derived from stem cells, *Methods in Enzymol.* 420:410-430.
72. Lian, X. and Palecek, S.P. (2012) Biomaterials for human pluripotent stem cells for therapeutic applications in Advances in Stem Cell Research, *Stem Cell Biology and Regenerative Medicine*, pp 29-48.
73. Baker, M. (2011) Stem cells in culture: defining the substrate, *Nature Methods* 8:293-297.
74. Fuchs, E. (2012) The impact of cell culture on stem cell research, *Cell Stem Cell* 10:640-641.
75. Xu, C. et al (2001) Feeder-free growth of undifferentiated embryonic stem cells, *Nature Biotechnol.* 19:971-974.
76. Ludwig, T.E. et al (2006) Feeder-independent culture of human embryonic stem cells, *Nature Methods* 3:637-646.
77. Yao, S. et al (2005) Long-term self-renewal and directed differentiation of human embryonic stem cells in chemically defined conditions, *PNAS* 103:6907-6912.
78. Ludwig, T.E. et al (2006) Derivation of human embryonic stem cells in defined conditions, *Nature Biotechnology* 24:185-187.
79. Li, Y. et al (2005) Expansion of human embryonic stem cells in defined serum-free medium devoid of animal-derived products, *Biotech. Bioengineering* 91:688-698.
80. Amit, M et al (2004) Feeder layer- and serum-free culture conditions of human embryonic stem cells, *Biol. Rep.* 70:837-845.
81. Lu, J. et al (2006) Defined culture conditions of human embryonic stem cells, *PNAS* 103:5688-5693.
82. Braam, S. R. et al (2008) Recombinant Vitronectin Is a Functionally Defined Substrate That Supports Human Embryonic Stem Cell Self-Renewal via α V β 5 Integrin, *Stem Cells* 26:225.
83. Meng, G. et al (2012) Synergistic effect of medium, matrix and exogenous factors on the adhesion and growth of human pluripotent stem cells under defined, xeno-free conditions, *Stem Cells and Dev.* 21:2036-2048.
84. Saxena, D. and Qian, S. (2009) Laminin/entactin complex: a feeder-free surface for culture of human embryonic stem cells, Corning Life Sciences - Application Note #464.
85. Rodin, S. et al (2011) Long-term self-renewal of human pluripotent stem cells on human recombinant laminin-511, *Nature Biotech* 28:611-615.
86. Li, Y.J., et al (2006) Hydrogels as artificial matrices for human embryonic stem cell self-renewal, *J Biomedical Materials Res Part A* 79A:1-5.
87. Villa-Diaz, L.G., et al (2010) Synthetic polymer coatings for long-term growth of human embryonic stem cells, *Nature Biotech* 28:581-583.
88. Brafman, D.A., et al (2010) Long-term human pluripotent stem cell self-renewal on synthetic polymer surfaces, *Biomaterials* 31:9135-9144.
89. Melkourian, Z., et al (2010) Synthetic peptide-acrylate surfaces for long-term self-renewal and cardiomyocyte differentiation of human embryonic stem cells, *Nature Biotech* 28:606-610.
90. Thompson, K., et al (2012) Corning PureCoat ECM Mimetic Cultureware Fibronectin Peptide: Novel Synthetic, Animal-free Surface for Culture of Human Bone Marrow-derived Mesenchymal Stem Cells, Corning Life Sciences - Application Note #494; Partridge, J., et al (2012) Corning PureCoat ECM Mimetic Cultureware Fibronectin Peptide: Synthetic and Animal-free Surface for Culture of Human Bone Marrow-derived Mesenchymal Stem Cells in StemPro[®] MSC SFM Xeno-free Medium, Corning Life Sciences - Application Note #496.
91. Tibbitt, M.W., et al (2009) Hydrogels as Extracellular Matrix Mimics for 3D Cell Culture, *Biotech and Bioeng* 103:655-663.
92. Seliktar, D. (2012) Designing Cell-Compatible Hydrogels for Biomedical Applications, *Science* 336:1124-1128.
93. Bhattacharyya, S., et al (2012) The voyage of stem cell toward differentiation: a brief overview, *Acta Biochim Biophys Sin* 44:463-475.
94. Arnautova, I., et al (2012) Basement Membrane Matrix (BME) has Multiple Uses with Stem Cells, *Stem Cell Rev and Rep* 8:163-169.
95. Itskovitz-Eldor, J., et al (2000) Differentiation of Human Embryonic Stem Cells into Embryoid Bodies Comprising the Three Embryonic Germ Layers, *Molec Med* 6:88-95.
96. Bratt-Leal, A.M., et al (2009) Engineering the Embryoid Body Microenvironment to Direct Embryonic Stem Cell Differentiation, *Biotechnol Prog* 25:43-51.
97. Hwang, N.S., et al (2008) Controlled differentiation of stem cells, *Adv Drug Deliv Rev* 60:199-214.
98. Ng, E.S., et al (2008) A protocol describing the use of a recombinant protein-based, animal-free medium (APEL) for human embryonic stem cell differentiation as spin embryoid bodies, *Nature Protocols* 3:768-776.
99. Lysashenko, N., et al (2011) Differential requirement for the dual functions of β -catenin in embryonic stem cell self-renewal and germ layer formation, *Nature Cell Biology* 13:753-761.
100. Zhou, L., et al (2011) Differentiation of Induced Pluripotent Stem Cells of Swine into Rod Photoreceptors and Their Integration into the Retina, *Stem Cells* 29:972-980.
101. Levenburg, S., et al (2002) Endothelial cell derived from human embryonic stem cells, *PNAS* 99:4391-4396.
102. Levenberg, S., et al (2003) Differentiation of human embryonic stem cells on three-dimensional polymer scaffolds, *PNAS* 100:12741-12746.
103. Escobedo-Lucca, C., et al (2012) Development of a Human Extracellular Matrix for Applications Related with Stem Cells and Tissue Engineering, *Stem Cell Rev and Rep* 8:170-183.
104. Philp, D.P., et al (2005) Complex Extracellular Matrices Promote Tissue-Specific Stem Cell Differentiation, *Stem Cells* 23:288-296.
105. Battista, S., et al (2005) The effect of matrix composition of 3D constructs on embryonic stem cell differentiation, *Biomaterials* 26:6194-6207.
106. Torres, J., et al (2012) Efficient Differentiation of Embryonic Stem Cells into Mesodermal Precursors by BMP, Retinoic Acid and Notch Signalling, *PLoS One* 7(4):e36405.
107. Tannenbaum, S.E. et al (2012) Derivation of Xeno-Free and GMP-Grade Human Embryonic Stem Cells – Platforms for Future Clinical Applications, *PLoS One* 7(6):e35325.
108. Engler, A.J., et al (2006) Matrix Elasticity Directs Stem Cell Lineage Specification, *Cell* 126:677-689.
109. Oh, S., et al (2009) Stem cell fate dictated solely by altered nanotube dimension, *PNAS* 106:2130-2135.
110. Du, J., et al (2011) Integrin activation and internalization on soft ECM as a mechanism of induction of stem cell differentiation by ECM elasticity, *PNAS* 108:9466-9471.
111. Nava, M.M., et al (2012) Controlling Self-Renewal and Differentiation of Stem Cells via Mechanical Cues, *J Biomed and Biotech* DOI:10.1155/2012/797410.
112. Lutolf, M.P., et al (2009) Designing materials to direct stem cell fate, *Nature* 462:433-441.
113. Vunjak-Novakovic, G. and Scadden, D.T. (2011) Biomimetic Platforms for Human Stem Cell Research, *Cell Stem Cell* 8:252-261.
114. Dalby, M.J. et al (2007) The control of human mesenchymal cell differentiation using nanoscale symmetry and disorder *Nature Materials* 6:997-1003.
115. Kshitzit, et al (2011) Micro- and nano-engineering for stem cell biology: the promise with a caution *Trends in Biotechnology* 29:399-408.
116. Kim, D.-H. (2012) Matrix nanotopography as a regulator of cell function *J. Cell Biol.* 197:351-360.
117. Kobel, S. and Lutolf, M.P. (2011) Biomaterials meet microfluidics: building the next generation of artificial niches *Curr. Opin. Biotechnology* 22:690-697.

REFERENCES

118. Chen, H.-F. et al (2007) Derivation, characterization and differentiation of human embryonic stem cells: comparing serum-containing versus serum-free media and evidence of germ cell differentiation *Human Reproduction* 22:567-577.
119. Rowland, T.J. et al (2010) Roles of integrins in human induced pluripotent stem cell growth on matrigel and vitronectin stem cells *Dev.* 19:1231-1240.
120. Turovets, N. et al (2012) Derivation of high-purity definitive endoderm from human parthenogenetic stem cells using an in vitro analog of the primitive streak, *Cell Transplantation* 21:217-234.
121. Kopper, O. and Benvenisty, N. (2012) Stepwise differentiation of human embryonic stem cells into early endoderm derivative and their molecular characterization, *Stem Cell Res.* 8:335-345.
122. Fujiwara, H. et al (2007) Regulation of mesodermal differentiation of mouse embryonic stem cells by basement membrane, *J. Biol. Chem.* 282:29701-29711.
123. Tada, S. et al (2005) Characterization of mesoderm: a diverging point of the definitive endoderm and mesoderm in embryonic stem cell differentiation culture, *Development* 132:4363-4374.
124. Grecht, S. et al (2007) Hyaluronic acid hydrogel for controller self-renewal and differentiation of human embryonic stem cells, *PNAS* 104:11298-11303.
125. Park, S.W. et al (2012) Efficient differentiation of human pluripotent stem cells into functional CD34+ progenitor cells by combined modulation of the MEK/ERK and BMP4 signaling pathways, *Blood* 16:5762-5772.
126. Carlson, A.L. et al (2012) Microfibrous substrate geometry as a critical trigger for the organization, self-renewal, and differentiation of human embryonic stem cells within synthetic 3-dimensional microenvironments, *FASEB J.* 26:3240-3251.
127. Uemura, M. et al (2010) Matrigel supports survival and neuronal differentiation of grafted embryonic stem cell-derived neural precursor cells, *J. Neuroscience Res.* 88:542-551.
128. Brannvall, K. et al (2007) Enhanced neuronal differentiation in a three-dimensional collagen-hyaluronan matrix, *Journal of Neuroscience Research* 85:2138-2146.
129. Itsykson, P. (2005) Derivation of neural precursors from human embryonic stem cells in the presence of noggin, *Mol. Cell. Neuroscience* 30:24-36.
130. Di Giorgio, F.P. et al (2008) Human embryonic stem cell-derived motor neurons are sensitive to the toxic effect of glial cells carrying an ALS-causing mutation, *Cell Stem Cell* 3:637-648.
131. Hutton, S.R. and Penvy, L. H. (2008) Isolation, culture, differentiation of progenitor cells from the central nervous system, *Cold Spring Harbor Protocols* 3(11):1-5.
132. Eiraku, M. et al (2008) Self-organized formation of polarized cortical tissues from ESCs and its active manipulation by extrinsic signals, *Cell Stem Cell* 3:519-532.
133. Eiraku, M. and Sasai, Y. (2012) Self-formation of layered neural structures in three-dimensional culture of ES cells, *Curr. Opin. Neurobiology* 22:768-777.
134. Wataya, T. et al (2008) Minimization of exogenous signals in ES cell culture induces rostral hypothalamic differentiation, *PNAS* 105:11796-11801.
135. Ma, Z. et al (2005) Potential of nanofiber matrix as tissue-engineering scaffolds, *Tissue Engineering* 11:101-109.
136. Li, X. et al (2012) Engineering neural stem cell fates with hydrogel design for central nervous system regeneration, *Progress in Polymer Science* 37(8):1105-1129.
137. Willerth, S.M. et al (2006) Optimization of fibrin scaffolds for differentiation of murine embryonic stem cells into neural lineage cells, *Biomaterials* 27:5990-6003.
138. Gaspard, N. et al (2008) An intrinsic mechanism of corticogenesis from embryonic stem cells, *Nature* 455:351-357.
139. Li, R. and Mather, J.P. (2008) Culture of pluripotent neural epithelial progenitor cells from E9 rat embryo, *Methods in Cell Biology* 86:227-240.
140. Bielby, R.C. et al (2004) In vitro differentiation and in vitro mineralization of osteogenic cells derived from human embryonic stem cells, *Tissue Engineering* 10:1518-1525.
141. Smith, L.A. et al (2009) The influence of three dimensional nanofibrous scaffolds on the osteogenic differentiation of embryonic stem cells, *Biomaterials* 10(13):2516-2522.
142. Garreta, E. et al (2006) Osteogenic differentiation of mouse embryonic stem cells and mouse embryonic fibroblasts in a three-dimensional self-assembling peptide scaffold, *Tissue Engineering* 12:2215-2227.
143. Chiang, P.-M and Wong, P.C. (2011) Differentiation of an embryonic stem cell to hemogenic endothelium by defined factors: essential role of bone morphogenetic protein 4, *Development* 138:2833-2843.
144. Wang, Y. et al (2005) Embryonic stem cell-derived hematopoietic stem cells, *PNAS* 102:19081-19086.
145. Liu, H. et al (2006) Effect of 3D scaffold and dynamic culture condition on the global gene expression profile of mouse embryonic stem cells, *Biomaterials* 27:5978-5989.
146. Kusuma, S. et al (2011) Derivation of endothelial cells from pluripotent stem cells in a feeder-free, two dimensional niche, *FASEB J.* 25:1043-12.
147. Joo, H.J. (2012) ROCK suppression promotes differentiation and expansion of endothelial cells from embryonic stem cell-derived Flk1+ mesodermal precursor cells, *Blood* 120:2733-2744.
148. Lee, S.-J. et al (2011) Robust vessel forming capability of endothelial cells which are differentiated from human pluripotent stem cells via a novel two-dimensional serum free differentiation system, *Circulation* 124:A16809.
149. Joo, H.-J. (2012) angiopoietin-1 promotes endothelial differentiation from embryonic stem cells and induced pluripotent stem cells, *Blood* 118:2094-2104.
150. Laflamme, M.A., et al (2007) Cardiomyocytes derived from human embryonic stem cells in pro-survival factors enhance function of infarcted rat hearts, *Nature Biotech* 25:1015-1024.
151. Kattman, S.J., et al (2011) Stage-Specific Optimization of Activin/Nodal and BMP Signaling Promotes Cardiac Differentiation of Mouse and Human Pluripotent Stem Cell Lines, *Cell Stem Cell* 8:228-240.
152. Ao, A., et al (2012) DMH1, a Novel BMP Small Molecule Inhibitor, Increases Cardiomyocyte Progenitors and Promotes Cardiac Differentiation in Mouse Embryonic Stem Cells, *PLoS One* 7(7):e41627.
153. Kehat, I., et al (2001) Human embryonic stem cells can differentiate into myocytes with structural and functional properties of cardiomyocytes, *J Clin Invest* 108:407-414.
154. Yang, L., et al (2008) Human cardiovascular progenitor cells develop from a KDR+ embryonic-stem-cell-derived population, *Nature* 453:524-528.
155. Guo, X.-M., et al (2012) Creation of Engineered Cardiac Tissue In Vitro from Mouse Embryonic Stem Cells, *Circulation* 113:2229-2237.
156. Flaim, C.J. et al (2008) Combinatorial signaling microenvironments for studying stem cell fate, *Stem Cells Dev.* 17:29-39.
157. Young, J.L. and Engler, A.J. (2011) Hydrogels with time-dependent material properties enhance cardiomyocyte differentiation in vitro, *Biomaterials* 32:1002-1009.
158. Mummery, C.L., et al (2012) Differentiation of Human Embryonic Stem Cells and Induced Pluripotent Stem Cells to Cardiomyocytes, *Circ Res* 111:344-358.
159. Carpenter, L., et al (2012) Efficient Differentiation of Human Induced Pluripotent Stem Cells Generates Cardiac Cells that Provide Protection Following Myocardial Infarction in the Rat, *Stem Cells and Dev* 21:977-986.
160. Zhang, J., et al (2012) Extracellular Matrix Promotes Highly Efficient Cardiac Differentiation of Human Pluripotent Stem Cells: The Matrix Sandwich Method, *Circ Res* 111:1125-1136.
161. Wang, D., et al (2007) A pure population of lung alveolar epithelial type II cells derived from human embryonic stem cells, *PNAS* 104:4449-4454.
162. Banerjee, E.R., et al (2012) Human Embryonic Stem Cells Differentiated to Lung Lineage-Specific Cells Ameliorate Pulmonary Fibrosis in a Xenograft Mouse Model, *PLoS One* 7(3):e33165.
163. Longmire, T.A., et al (2012) Efficient Derivation of Purified Lung and Thyroid Progenitors from Embryonic Stem Cells, *Cell Stem Cell* 10:398-411.
164. Mou, H., et al (2012) Generation of Multipotent Lung and Airway Progenitors from Mouse ESCs and Patient-Specific Cystic Fibrosis iPSCs, *Cell Stem Cell* 10:385-397.
165. Gong, J.O. et al (2008) Effects of extracellular matrix and neighboring cells on induction of human embryonic stem cells into retinal or retinal pigment epithelial progenitors *Exp. Eye Res* 86:957-965.
166. Lambda, D.A., et al (2010) Generation, Purification and Transplantation of Photoreceptors Derived from Human Induced Pluripotent Stem Cells, *PLoS One* 5(1):e8763.
167. Zahabi, A., et al (2012) A New Efficient Protocol for Directed Differentiation of Retinal Pigmented Epithelial Cells from Normal and Retinal Disease Induced Pluripotent Stem Cells, *Stem Cells and Dev* 21:2262-2272.
168. Nistor, G. (2010) Three-dimensional early retinal progenitor 3D tissue constructs derived from human embryonic stem cells, *J Neurosci Methods* 190:63-70.
169. Eiraku, M., et al (2011) Self-organizing optic-cup morphogenesis in three-dimensional culture, *Nature* 472:51-56.
170. Buchholz, D.E., et al (2009) Derivation of Functional Retinal Pigmented Epithelium from Induced Pluripotent Stem Cells, *Stem Cells* 27:2427-2434.
171. Rowland, T.J., et al (2012) Differentiation of human pluripotent stem cells to retinal pigmented epithelium in defined conditions using purified extracellular matrix proteins, *J Tissue Eng and Regen Med*, DOI:10.1002/term.1458.

REFERENCES

172. Krohne, T.U., et al (2012) Generation of Retinal Pigment Epithelial Cells from Small Molecules and OCT4 Reprogrammed Human Induced Pluripotent Stem Cells, *Stem Cells Translational Medicine* 1:96-109.
173. Arufe, M.C., et al (2006) Directed Differentiation of Mouse Embryonic Stem Cells into Thyroid Follicular Cells, *Endocrinology* 147:3007-3015.
174. Antonica, F., et al (2012) Generation of functional thyroid from embryonic stem cells, *Nature* 49:66-71.
175. Suga, H., et al (2011) Self-formation of functional adeno-hypophysis in three-dimensional culture, *Nature* 480:57-62.
176. Zhou, W.-L., et al (2012) Stem cell differentiation and human liver disease, *World J Gastroenterol* 18:2018-2025.
177. Zhao, D., et al (2009) Derivation and Characterization of Hepatic Progenitor Cells from Human Embryonic Stem Cells, *PLoS One* 4(7):e6468.
178. Ishii, T., et al (2008) Effects of extracellular matrices and growth factors on the hepatic differentiation of human embryonic stem cells, *Am J Physiol Gastrointest Liver Physiol* 295:G313-G321.
179. Roelandt, P., et al (2010) Human Embryonic and Rat Adult Stem Cells with Primitive Endoderm-Like Phenotype Can Be Fated to Definitive Endoderm, and Finally Hepatocyte Like Cells, *PLoS One* 5(8):e12101.
180. Sullivan, P., et al (2010) Generation of Functional Human Hepatic Endoderm from Human Induced Pluripotent Stem Cells, *Hepatology* 51:329-335.
181. Si-Tayeb, K., et al (2010) Highly Efficient Generation of Human Hepatocyte-Like Cells from Induced Pluripotent Stem Cells, *Hepatology* 51:297-305.
182. Yamada, T., et al (2002) In Vitro Differentiation of Embryonic Stem Cells into Hepatocyte Like Cells Identified by Cellular Uptake of Indocyanine Green, *Stem Cells* 20:146-154.
183. Mandal, A., et al (2012) Stage specific differentiation of human embryonic stem cells into hepatocyte-like cells using conditioned medium from a human hepatoma cell line, *Stem Cell Studies* 2:e2.
184. Shirasawa, S., et al (2011) A novel stepwise differentiation of functional pancreatic exocrine cells from embryonic stem cells *Stem Cells Dev.* 20:1071-1079.
185. Chen, S., et al (2009) A small molecule that directs differentiation of human ESCs into the pancreatic lineage *Nature Chemical Biology* 5:258-265.
186. Zhang, D., et al (2009) Highly efficient differentiation of human ES cells and iPSC cells into mature pancreatic insulin-producing cells *Cell Research* 19:429-438.
187. Tanaka, K. et al (2004) Chondrogenic differentiation of murine embryonic stem cells: Effects of culture conditions and dexamethasone, *J. Cellular Biochemistry* 93:454-462.
188. Hwang, N.S., et al (2006) Effects of three-dimensional culture and growth factors on the chondrogenic differentiation of murine embryonic stem cells, *Stem Cells* 24:284-291.
189. Sato, T. et al (2009) Single Lgr5 stem cells build crypt-villus structures in vitro without a mesenchymal niche, *Nature* 459:262-265.
190. Barker, N. et al (2010) Lgr5+ve stem cells drive self-renewal in the stomach and build long-lived gastric units in vitro, *Cell Stem Cell* 6:25-36.
191. Sato, Y. et al (2011) Long term expansion of epithelial organoids from human colon, adenoma, adenocarcinoma, and Barrett's epithelium, *Gastroenterology* 141:1762-1772.
192. Vermeulen, L. et al (2008) Single cell cloning of colon cancer stem cells reveals a multilineage differentiation capacity, *PNAS* 105:13427-13432.
193. Van Dussen, K.L. (2012) Notch signaling modulated proliferation and differentiation of intestinal crypt base columnar stem cells, *Development* 139:488-497.
194. Snippert, H.J. et al (2012) Intestinal crypt homeostasis results from neutral competition between symmetrically dividing Lgr5 stem cells, *Cell* 143:134-144.
195. Ootani, A. et al (2009) Sustained in vitro intestinal epithelial culture within a Wnt-dependent stem cell niche, *Nature Medicine* 17:701-706.
196. Jung, P. et al (2011) Isolation and in vitro expansion of human colonic stem cells, *Nature Medicine* 17:1225-1227.
197. Yui, S. et al (2012) Functional engraftment of colon epithelium expanded in vitro from a single adult Lgr5+ stem cell, *Nature Medicine* 18:618-623.
198. Shackleton, M. et al (2006) Generation of a functional mammary gland from a single stem cell, *Nature* 439:84-88.
199. Stingl, J. et al (2006) Purification and unique properties of mammary epithelial stem cells *Nature* 439:993-997.
200. Guo, W. et al (2012) Slug and Sox9 cooperatively determine the mammary stem cell state *Cell* 148:1015-1028.
201. Blanpain, C. et al (2004) Self renewal, multipotency, and the existence of two populations within an epithelial stem cell niche, *Cell* 118:635-648.
202. Connelly, J.T. et al (2010) Actin and serum response factor transduce physical cues from the microenvironment to regulate epidermal stem cell fate decisions, *Nature Cell Biology* 12:711-718.
203. Yu, H. et al (2006) Isolation of a novel population of multipotent adult stem cells from human hair follicles, *Am. J. Pathol.* 168:1879-1888.
204. Gao, R. et al (2003) Characterization of endocrine progenitor cells and critical factors for their differentiation in human adult pancreatic cell culture *Diabetes* 52:2007-2015.
205. Yebra, M. et al (2011) Endothelium-derived netrin-4 supports pancreatic epithelial cell adhesion and differentiation through integrins $\alpha 2\beta 1$ and $\alpha 3\beta 1$, *PLoS ONE* 6:e22750.
206. Seaburg, R.M. et al (2004) Clonal identification of multipotent precursors from adult mouse pancreas that generate neural and pancreatic lineages, *Nature Biotechnology* 22:1115-1124.
207. Messina, E. et al (2004) Isolation and expansion of adult cardiac stem cells from human and murine heart, *Circulation Res.* 95:911-921.
208. Bayomy, A.F. et al (2012) Regeneration in heart disease-is ECM the key?, *Life Sciences* 91:823-827.
209. Chang, Y.-J. et al (2012) Endothelium differentiation and regeneration from human adipose-derived stem cell by sheer stress, *FASEB J.* 26:639.11.
210. Cao, Y. et al (2005) Human adipose tissue-derived stem cells differentiate into endothelial cells in vitro and improve postnatal neovascularization in vivo, *Biochem. Biophys. Res. Comm.* 332:370-379.
211. Yang, L. et al (2002) In vitro trans-differentiation of adult hepatic stem cells into pancreatic endocrine hormone-producing cells, *PNAS* 99:8078-8083.
212. Herrera, M.B. et al (2006) Isolation and characterization of a stem cell population from adult human liver, *Stem Cells* 24:2840-2850.
213. Lucendo-Villarin, B. et al (2012) Maintaining hepatic stem cell gene expression on biological and synthetic substrata, *BioResearch Open Access* 1(1):50-53.
214. Bender Kim, C.F. et al (2005) Identification of bronchioalveolar stem cells in normal lung and lung cancer, *Cell* 121:823-835.
215. Okumura, K. et al (2012) Capability of tissue stem cells to organize into salivary glands, *Stem Cells International* ID#502136.
216. Phillips, M.J. and Otteson, D.C. (2011) Differential expression of neuronal genes in Muller glia in two- and three-dimensional culture, *IOVS* 52:1439-1449.
217. Demontis, G.C. et al (2012) Functional and molecular characterization of rod-like cells from retinal stem cells derived from the adult ciliary epithelium, *PLoS ONE* 7(3):23338.
218. Loov, C. et al (2012) Neutralization of LINGO-1 during in vitro differentiation of neural stem cells results in proliferation of immature neurons, *PLoS ONE* 7(1):e29771.
219. In-Hyeup Park, T. et al (2012) Adult human brain neural progenitor cells (NPCs) and fibroblast-like cells have similar properties in vitro but only NPCs differentiate into neurons, *PLoS ONE* 7(6):e37742.
220. Banerjee, A. et al (2009) The influence of hydrogel modulus on the proliferation and differentiation of encapsulated neural stem cells, *Biomaterials* 30:4695-4699.
221. Georges, P.C. et al (2006) Matrices with compliance comparable to that of brain tissue select neuronal over glial growth in mixed cortical cultures, *Biophysical Journal* 90:3012-3018.
222. Qian, L. and Saltzman, W.M. (2004) Improving the expansion and neuronal differentiation of mesenchymal stem cells through culture surface modification, *Biomaterials* 25:1331-1337.
223. Silva, G. et al (2004) Selective differentiation of neural progenitor cells by high epitope density nanofibers, *Science* 303:1352-1355.
224. Gelain, F. et al (2006) Designer self-assembling peptide nanofiber scaffolds for adult mouse neural stem cell 3-dimensional cultures, *PLoS ONE* 1:e119.
225. Thonhoff, J.R. et al (2008) Compatibility of human fetal neural stem cells with hydrogel biomaterials in vitro, *Brain Res.* 1187:42-51.
226. Ortinou, S. et al (2010) Effect of 3D-scaffold formation on differentiation and survival in human neural progenitor cells, *Biomedical Engineering Online* 9:70.
227. Cunha, C. et al (2011) 3D culture of adult mouse neural stem cells within functionalized self-assembling peptide scaffolds, *Int. J. Nanomedicine* 6:943-955.
228. Liedmann, A. et al (2012) Differentiation of human neural progenitor cells in functionalized hydrogel matrices, *BioResearch Open Access* 1(1):16-24.

REFERENCES

229. Saha, K. et al (2008) Substrate modulus directs neural stem cell behavior, *Biophysical J.* 95:4426-4438.
230. Lee, J.H. et al (2011) Collagen gel three-dimensional matrices combined with adhesive proteins stimulate neuronal differentiation of mesenchymal stem cells, *J. R. Soc. Interface* 8:998-1010.
231. Tan, G. et al (2010) Differential effect of myocardial matrix and integrins on cardiac differentiation of human mesenchymal stem cells, *Differentiation* 79:260271.
232. Flanagan, L.A. et al (2002) Neurite branching on deformable substrates, *Neuroreport* 13:2411-2415.
233. Tay, C.Y. et al (2010) Micropatterned matrix directs differentiation of human mesenchymal stem cells towards myocardial lineage, *Exp. Cell Res.* 316:1159-1168.
234. Santiago, J.A. et al (2009) Heterogeneous differentiation of human mesenchymal stem cells in response to extended culture in extracellular matrices, *Tissue Engineering: Part A* 15:3911-3922.
235. Singh, P. and Schwarzbauer, J.E. (2012) Fibronectin and stem cell differentiation – lessons from chondrogenesis, *J. Cell Sci.* 125:3703-3712.
236. Erickson, I.E. et al (2009) Differential maturation and structure-function relationships in mesenchymal stem cell- and chondrocyte-seeded hydrogels, *Tissue Engineering* 15:1041-1052.
237. Hamada, K. et al (2007) Spatial distribution of mineralized bone matrix produced by marrow mesenchymal stem cells in self-assembling peptide hydrogel scaffold, *Wiley InterScience DOI:10.1002/jbm.a.31439*.
238. Ozeki, M. et al (2010) Differentiation of bone marrow stromal cells into osteoblasts in a self-assembling peptide hydrogel: in vitro and in vivo studies *J. Biomaterials Applications* 25:663-684.
239. Trappman, B. et al (2012) Extracellular-matrix tethering regulates stem-cell fate, *Nature Materials* 11:642-649.
240. Firth, J.E. et al (2012) Tailored integrin-extracellular matrix interactions to direct human mesenchymal stem cell differentiation, *Stem Cells Dev.* 21:2442-2456.
241. Jing, D. et al (2010) Hematopoietic stem cells in co-culture with mesenchymal stromal cells-modeling the niche compartments in vitro, *Haematologica* 95:542-550.
242. Sharma, M.B. et al (2011) Mimicking the functional hematopoietic stem cell niche in vitro: recapitulation of marrow physiology by hydrogel-based three-dimensional cultures of mesenchymal stromal cells, *Haematologica* 97:652-660.
243. Jiang, Y. et al (2002) Pluripotency of mesenchymal stem cells derived from adult marrow, *Nature* 418:41-49.
244. Allen, P. et al (2011) Type 1 collagen, fibrin, and puramatrix matrices provide permissive environments for human endothelial and mesenchymal progenitor cells to form neovascular networks, *J. Tissue Eng. Reg. Medicine* 5:e74-e86.
245. Zhang, S. et al (1995) Self-complementary oligopeptide matrices support mammalian cell attachment, *Biomaterials* 16:1385-1393.
246. Semino, C.E. et al (2008) Self-assembling peptides: from bio-inspired materials to bone regeneration, *J. Dental Research* 87:606-616.
247. McNamara, L.E. et al (2010) Nanotopographical control of stem cell differentiation, *J. Tissue Engineering* ID# 120623.
248. Zhong, Y. and Bellamkonda, R.V. (2008) Biomaterials for the central nervous system, *J. R. Soc. Interface* 5:957-975.
249. Jensen, T. et al (2012) A rapid de-cellularization protocol supports embryonic stem cell differentiation in vitro following implantation, *Tissue Engineering: Part C* 18:632-646.
250. Liu, J. et al (2012) Generation, characterization, and potential therapeutic applications of cardiomyocytes from various stem cells, *Stem Cells and Dev.* 21:2095-2110.
251. Kosovsky, M. (2012) Culture and assay systems used for the study of cancer stem cells, Corning Life Sciences.

**Corning acquired the Cell-Tak™, Matrigel®, and PureCoat™ brands.
For information, visit www.corning.com/discoverylabware.**

For Research Use Only. Not for use in diagnostic or therapeutic procedures.
For a listing of trademarks, visit us at www.corning.com/lifesciences/trademarks.
All other trademarks in this document are the property of their respective owners.
Corning Incorporated, One Riverfront Plaza, Corning, NY 14831-0001

CORNING

Corning Incorporated
Life Sciences

836 North St.
Building 300, Suite 3401
Tewksbury, MA 01876
t 800.492.1110
t 978.442.2200
f 978.442.2476

www.corning.com/lifesciences